Intestinal Bacterial Flora that Compete on the Haem Precursor Iron Fumarate in Iron Deficiency Anemia Cases

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ABSTRACT

Aims: The study focused on finding if there is any possible relation between the intestinal bacterial population quantitative and qualitative and the deficiency of the most important iron compounds as haem precursors.

Methodology and Results: Blood complete picture and stool analyses were done to 750 volunteer cases whom were asked for these analyses by their physicians. Analyses proved that 560 cases representing 75.2 % were anemic as the RBC(s) based on counts of the total studied cases of less than 263 x 106 and the haemoglobin amount ranged between 7.2 and 11.3 g/dl, while the remainder 24.8 % of the volunteer sample was not anemic. A high male/female ratio of anemic cases, 1.27 was also documented. Considering that all the studied stool samples should be completely free from any parasites or any other anemia-related diseases was a priority. Bacteriological analysis of stool samples of the anemic cases resulted in the detection of high counts of total viable bacteria, exceeded 42 x 106 cfu/g, while it was never more than 26 x 106 cfu/g and decreased to 4 x 105 cfu/g in many cases in this study. Identifying of the 361 bacterial isolates, were found to belong to 12 genera and 19 species, 6 of them; Pseudomonas putrefaciens, Micrococcus luteus, Erysipelothrix rhusiopathiae, Bacillus megaterium, Bacillus pumilus and Bacillus coagulans , were found and in high counts in the stool samples of only anemic cases. The ability of these isolates to compete for iron compounds such as ferrous fumarate alone or with glucose and phytate as activators or inhibitors to these abilities was investigated. Results proved 11 species out of the 19 identified species are capable to use and compete on ferrous fumarate as a haem precursor. Sensitivity test for the representatives of the 19 species and 6 of the most commonly used antibiotics in the Egyptian pharmacy, using standard disc method, revealed variable susceptibilities of almost all of them to more than one of the studied antibiotics, except Corynebacterium equatumum, which was found very resistant to two antibiotics; colistin sulfate and erythromycin.

Conclusion, significance and impact of study: The study finally concluded the strong role of intestinal bacterial counts and types as competitors on the haem precursor iron-containing compounds like ferrous fumarate.

Keywords: Intestinal Bacterial Flora, Haem Precursor, Iron Fumarate, Iron Deficiency Anemia.

INTRODUCTION

Anemia is one of the more common blood disorders. Anemia can be defined as having less than the normal number of red blood cells or less hemoglobin than normal in the blood. There are several types of anemia and the most common type of anemia is iron deficiency anemia that happens when there is a deficiency of iron in the body. Iron deficiency anemia occurs when a person loses blood from problems such as heavy periods, ulcers, colon polyps, or colon cancer. In addition, a diet that does not have enough iron can also cause iron deficiency anemia (Hallberg, 1982; Layrisse et al., 2000). Iron is essential to all microorganisms. To obtain iron from the very low concentrations present in their environment, microorganisms have developed sophisticated mechanisms such as the siderophore system (Bagg and Neilands, 1987; Crichton and Ward, 1992). Anemia is seen in the setting of infectious, inflammatory, and neoplastic diseases. It results, from changes in the intracellular metabolism of iron. Alterations of iron physiology seen in many clinical circumstances make excess iron available to microorganisms, thus enhancing their pathogenicity. Nearly all groups of potentially pathogenic bacteria thus far examined require iron for activation of ribonucleotide reductase, aconitase and various enzymes involved in electron transfer and oxygen metabolism (Bullen et al., 1978; Byers, 1987; Griffiths, 1987; 1991; Martinez et al., 1990; Braun et al., 1998).

The intestinal bacterial flora in patients with anemia has long attracted the interest of investigators, but few recent studies have been reported. The present investigation was undertaken to reappraise the significance of the bacterial flora of the small intestine and to correlate bacterial growth with intestinal absorption in patients with

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is iron deficiency anemia. The implication of intestinal bacteria in the pathogenesis of anemia began with the intestinal-toxin theories of William Hunter in 1900. There followed a long series of investigations of the gastrointestinal flora in anemia with successful attempts to demonstrate excesses of specific types of bacteria in this disease. Davidson (1928) studied the bacterial flora of the stomach and colon and found marked quantitative increases of microorganisms in these areas in patients with anemia. Although the upper gastrointestinal flora was frequently colonic in type, no particular organism was consistently present. Other investigators have noted a proliferation of fecal organisms in the upper gastrointestinal tract of patients with anemia (Conrad and Crosby, 1963; William et al., 1964).

The present study focused on determining the quantitative and qualitative variation of the intestinal bacterial population in relation with the iron deficiency anemia features in Egypt. Also the study investigated the possibility of prescribing antibacterial agents along with other anemia-treating drugs.

**MATERIALS AND METHODS**

**Patients**

A well-taken history from somebody close to the patient, as well as from the actual patient, particularly when there is doubt as to whether he is capable, or willing to give full details of diet, drug ingestion, was carefully and accurately recorded for every single case. Random survey of 750 volunteer cases in order to determine anemic cases and recorded for every single case. Random survey of 750 volunteer cases in order to determine anemic cases and healthy non-anemic cases comprised 75.2 % of the total sample size were suffering from obvious anemia symptoms, parasite- free and had no clearly-defined suspected cause of anemia. Healthy non-anemic control 107 volunteers were as fully as the anemic

**Measurements**

Whole blood specimens for haematology laboratory were collected by venepuncture technique (NCCLS 1992a). The syringe and needle method were followed as it is preferred when difficulty in drawing the specimen is anticipated, e.g. patients with fragile or hardened vein walls. Gentle inversion, five to ten times of the tube containing part of the blood sample and the anticoagulant, trisodium citrate (Na₃C₆H₅O₇) recommended in a 1:4 anticoagulant/blood ratio for determining the erythrocyte sedimentation rate (ESR) (ICSH, 1977). EDTA (C₁₀H₁₈N₂O₈) was added to the rest of the sample at 3.7-5.4 µmol/mL for performing other blood analyses including; haemoglobin value, red blood cell count, white blood cell count, mean corpuscular volume, mean corpuscular haemoglobin and mean corpuscular haemoglobin concentration (Lewis and Koepeke, 1995).

**Microbiological Analysis**

Two stool samples volume of at least 1 teaspoonful (5 mL) was collected in clean, waxed cardboard or plastic container and another in sterile glass vial for bacteriological studies and delivered to the laboratory with no longer than one hour for physical and microbiological examinations. The first was microscopically examined for parasites. One gram of the second was suspended in 100 mL sterile saline, serially diluted and 1 mL of the dilutions was cultivated on nutrient agar plates using the pour plate method. TVB counts cfu/mL were the recorded after 24 h incubation at 37 °C. Macro and micro-morphological descriptions as well as complete identification of the bacterial isolates following the biochemical charts (Holt, 1994) were achieved. Sensitivity test for all isolates was performed against six antibiotics namely; colistin sulphate, streptomycin, velosef, chloramphenicol, ampicillin and erythromycin using standard antibiotic discs (Murray et al., 1995).

**Utilization of Haem Precursor Compounds**

For the detection and estimation of bacterial isolates that compete for fumarate as a haem precursor, pure bacterial cultures were grown on nutrient agar for 24 h at 37 °C, then harvested and suspended in Davis minimal broth medium. Growth was measured and equally adjusted for all cultures absorbance spectrophotometrically at A₆₈₀. A set of four tubes was prepared as follows for each bacterial isolate; the first contained 1mL bacterial suspension and 4 mL of the minimal broth, the second was the same as the first supplemented with ferrous fumarate 1 µg/mL, the third was the same as the second plus phytic acid 7.3 µg/mL and the fourth was the same as the second plus glucose 7.3 µg/mL. In a shaker incubator, all tube sets were incubated at 37 °C for 48 h and the absorbance was read again at A₆₀₀. A standard curve was plotted and the extent of growth was directly determined from its absorbance.

**RESULTS**

Volunteer sample size covered in this study started with 750 cases. Non anemic cases (186) comprised 24.8 % of the total sample size, 9 % of these non-anemic cases were infected with parasites. Anemic cases (560) comprised 75.2 % of the total sample size were suffering from obvious anemia symptoms, parasite- free and had no clearly-defined suspected cause of anemia. Healthy non-anemic control 107 volunteers were as fully as the anemic
cases. Results of complete blood picture and parasitological stool analyses were the tools upon which samples were described and the study was designed. The sample included 448 females and 112 males with haemoglobin value (Hb), ranged from 7.2-11.7 for females and 11.3 for males, while control (Hb) values were above 12 up to 17.4 represented by 36 females and 71 males volunteers. Counts of RBC(s) ranged from 26 x 10^6 to 707 x 10^6. The erythrocyte sedimentation rate (ESR) ranged from 4-110 after 1 h and from 11 to 110 after 2 h. White blood cell counts ranged between 2625 and 11650. Other blood picture components such as P.C.V, M.C.V, M.C.H and M.C.H.C were examined and the results were not published.

Enumeration of bacteria in stool samples of the studied anemic cases showed high total viable bacterial counts (TVB) counts and great variations from sample to sample. A minimum of 13 x 10^6 cfu/g and a maximum of 42 x 10^8 cfu/g were recorded for anemic cases, both from females, while in the male sample TVB count was 75 x 10^6 cfu/g. The counts for control volunteer samples started from 4 x 10^6 (a female case) up to 26 x 10^6 (a male case). Bacterial isolates from stool samples (361 isolates) were identified and confirmed to belong to 12 genera and 19 species. Neisseria mucosa (190 isolates) comprised the highest percentage (52.6 %) of the total isolates as well as the highest distribution rate amongst all cases including control volunteers. In the second place came P. pseudocaligenes and P. putrefaciens (45 isolates) and they detected only in anemic cases. In addition to P. pseudocaligenes. More six species belong to four genera were detected only in stool samples of anemic cases namely, B. pumilus, B. megaterium, B. coagulans, E. rhusiopathiae, M. luteus, and A. viridans. Species those detected both in anemic and non-anemic stool specimens were dominated by N. mucosa with other eleven species namely; L. monocytogenes, S. aureus, B. cereus, B. subtilis, P. mirabilis, A. punctata, A. calcoaceticus, M. rosus, C. equatum and C. pseudotuberculosis.

Representatives of the identified species (19 isolates) were chosen to perform sensitivity test against six commonly used antibiotics. Erythrocin suppressed the growth of N. mucosa, L monocytogenes, M. luteus, P. pseudocaligenes, B. pumilus and E. rhusiopathiae. Streptomycin inhibited the growth of B. coagulans, B. megaterium, B. cereus, A. viridans, C. pseudotuberculosis, P. putrefaciens and M. luteus. Chloramphenical suppressed the growth S. aureus, A. punctate, M. roseus, M. luteus, P. mirabilis, A. calcoaceticus, B. megaterium and A. viridans. These three antibiotics seemed to be broad spectrum and have a quite wide range against most of the studied bacterial isolates and showed clear zones from 17 to 25 mm in diameter. The N. mucosa isolate was sensitive to all the six antibiotics and was highly susceptible to chloramphenicol, ampicillin and erythrocin having clear zone diameters of 10, 11 and 12 mm respectively. L. monocytogenes was more or less equally affected by all the studied six antibiotics. M. luteus showed higher susceptibility to all the studied antibiotics than M. roseus. P. pseudocaligenes showed high sensitivity to velosof and erythrocin, while P. putrefaciens strongly affected by streptomycin and ampicillin. Genus Bacillus showed variable susceptibility profiles. The tested five species were sensitive to all the studied antibiotics with high tendency of B. subtilis to streptomycin and velosof, B. pumilus to erythrocin, B. megaterium and B. cereus to chloramphenicol and B. coagulans to streptomycin. C. equatum was greatly affected by velosof and chloramphenicol, while showed complete resistance to colistin sulfate and erythrocin.

All the identified nineteen bacterial species isolated from stool samples of anemic cases and control volunteers were examined for their capability in using iron in the form of ferrous feumarate (FeFu) alone and in combination with phytate (ph) and or glucose (G). Three types of controls were necessary one without supplements, a second with only glucose (G) and a third with only phytate (ph). All the investigated isolates representing the 19 identified species took up iron in the form of (FeFu) but 10 of them namely; N. mucosa, A. punctata, M. roseus, M. luteus, P. mirabilis, P. putrefaciens, C. equatum, B. pumilus, B. megaterium B. coagulans, and E. rhusiopathiae consumed much more iron in the form of (FeFu) than the representative isolates of the other 9 species and gave relatively high absorbance readings ranged between 0.14-0.2 (Table 1). They showed enhanced usage of (FeFu) when combined with (ph) and with (G). The other 9 isolates showed variable responses in which, for example, (ph) and (G) acted as growth limiting factors as the bacterial growth solely on them was greater than in combinations with (FeFu).

Genus of Neisseria was presented by only one species; mucosa. It showed very little growth in the presence of (FeFu) alone and (G) alone, while the growth was very heavy in a combination of them both. Growth in (ph) alone was as the same as in a combination of (FeFu+ph). Genus Staphylococcus was also presented by only one species; aureus. The resulted growth rate was the same in (G) alone and (FeFu+G). Same results were recorded for (ph) alone and for (FeFu+ph). The four growth rate readings were high if compared with the very low reading of growth rate with (FeFu) alone. Species monocytogenes the only representative for genus Listeria, showed scanty growth in (FeFu) alone and in (FeFu+G) while it gave rich growth with (G) alone. The growth in (ph) alone and with (FeFu+ph) did not show any difference. A big difference was recorded between the growth rate of A. punctata in the presence of a combination of (FeFu+G), in one hand and the growth in (FeFu) alone or in (G) alone on the other hand, the former was much higher. No obvious difference was detected between growths in (ph) alone and in (FeFu+ph). A little bit better growth was observed in (FeFu+G) then in (G) alone or (FeFu) alone for the only species; mirabilis that presented genus Proteus. No growth differences were recorded in (FeFu+ph) than in (ph) alone.
Table 1: Growth after 48 h of bacterial isolates on non supplemented minimal broth medium as control and on minimal broth medium supplemented with ferrous fumarate (FeFu), glucose (G), ferrous fumarate + glucose (FeFu+G), phytate (ph) and with ferrous fumarate + phytate (FeFu+ph).

<table>
<thead>
<tr>
<th>Bacterial isolates</th>
<th>Control</th>
<th>FeFu</th>
<th>G</th>
<th>FeFu+G</th>
<th>ph</th>
<th>FeFu+ph</th>
</tr>
</thead>
<tbody>
<tr>
<td>A. calcoaceticus</td>
<td>0.012</td>
<td>0.076</td>
<td>0.087</td>
<td>0.118</td>
<td>0.152</td>
<td>0.019</td>
</tr>
<tr>
<td>A. viridans</td>
<td>0.01</td>
<td>0.007</td>
<td>0.049</td>
<td>0.114</td>
<td>0.129</td>
<td>0.13</td>
</tr>
<tr>
<td>A. punctata</td>
<td>0.01</td>
<td>0.002</td>
<td>0.012</td>
<td>0.119</td>
<td>0.112</td>
<td>0.126</td>
</tr>
<tr>
<td>B. subtilis</td>
<td>0.035</td>
<td>0.071</td>
<td>0.099</td>
<td>0.076</td>
<td>0.155</td>
<td>0.066</td>
</tr>
<tr>
<td>B. coagulans</td>
<td>0.01</td>
<td>0.006</td>
<td>0.011</td>
<td>0.15</td>
<td>0.185</td>
<td>0.125</td>
</tr>
<tr>
<td>B. megaterium</td>
<td>0.023</td>
<td>0.008</td>
<td>0.073</td>
<td>0.091</td>
<td>0.077</td>
<td>0.092</td>
</tr>
<tr>
<td>B. pumilus</td>
<td>0.022</td>
<td>0.007</td>
<td>0.098</td>
<td>0.181</td>
<td>0.084</td>
<td>0.021</td>
</tr>
<tr>
<td>B. cereus</td>
<td>0.016</td>
<td>0.015</td>
<td>0.011</td>
<td>0.026</td>
<td>0.024</td>
<td>0.011</td>
</tr>
<tr>
<td>C. equatum</td>
<td>0.098</td>
<td>0.01</td>
<td>0.145</td>
<td>0.153</td>
<td>0.111</td>
<td>0.018</td>
</tr>
<tr>
<td>C. pseudotuberculosis</td>
<td>0.01</td>
<td>0.01</td>
<td>0.104</td>
<td>0.122</td>
<td>0.113</td>
<td>0.136</td>
</tr>
<tr>
<td>E. rhusiopathiae</td>
<td>0.018</td>
<td>0.02</td>
<td>0.115</td>
<td>0.131</td>
<td>0.12</td>
<td>0.134</td>
</tr>
<tr>
<td>L. monocytogenes</td>
<td>0.023</td>
<td>0.011</td>
<td>0.164</td>
<td>0.011</td>
<td>0.103</td>
<td>0.106</td>
</tr>
<tr>
<td>M. leteus</td>
<td>0.017</td>
<td>0.017</td>
<td>0.084</td>
<td>0.145</td>
<td>0.11</td>
<td>0.125</td>
</tr>
<tr>
<td>M. roseus</td>
<td>0.016</td>
<td>0.016</td>
<td>0.157</td>
<td>0.136</td>
<td>0.149</td>
<td>0.163</td>
</tr>
<tr>
<td>N. mucosa</td>
<td>0.017</td>
<td>0.015</td>
<td>0.025</td>
<td>0.141</td>
<td>0.127</td>
<td>0.0134</td>
</tr>
<tr>
<td>P. putrefaciens</td>
<td>0.01</td>
<td>0.008</td>
<td>0.105</td>
<td>0.13</td>
<td>0.116</td>
<td>0.152</td>
</tr>
<tr>
<td>P. pseudoalcaligenes</td>
<td>0.019</td>
<td>0.011</td>
<td>0.097</td>
<td>0.113</td>
<td>0.098</td>
<td>0.015</td>
</tr>
<tr>
<td>P. mirabilis</td>
<td>0.017</td>
<td>0.018</td>
<td>0.102</td>
<td>0.135</td>
<td>0.119</td>
<td>0.125</td>
</tr>
<tr>
<td>S. aureus</td>
<td>0.018</td>
<td>0.014</td>
<td>0.118</td>
<td>0.12</td>
<td>0.1</td>
<td>0.115</td>
</tr>
</tbody>
</table>

Genus *Pseudomonas* had two representative species; *pseudoalcaligenes* and *putrefaciens*. Scanty growth was noticed for *P. pseudoalcaligenes* in (FeFu+ph) while growth was high in (G) alone, (FeFu+G) and (ph) alone. For *P. putrefaciens* growth rate in combinations of (FeFu+G) and (FeFu+ph) was higher than in (G) or (ph) alone. *A. calcoaceticus* gave slightly higher growth in (FeFu+G) than in (G) alone, but gave very poor growth in (FeFu+ph) if compared with the growth in (ph) alone. Genus *Corynebacterium* had two representative species in the present work; *equatum* and *pseudotuberculosis*. Growth of both in (FeFu) was much lower than in the control. The rate of growth increased, almost the same amount of increase, in (G), (ph) and in (FeFu+G) than in the control. *A. viridans* was the only species that gave negative growth reading (growth inhibited) with (FeFu) alone, high reading with (G) alone and higher readings with (FeFu+G), (FeFu+ph) and (ph). *E. rhusiopathiae* had nearly no changes in growth figures with different treatments.

**DISCUSSION**

None of the identified bacterial species in healthy control samples could be recognized as characterizing species for only non-anemic people, as they all were recorded in the studied anemic cases. The presence of certain bacterial species both in anemic and non-anemic control samples was observed, but the counts of each was higher in anemic than in non-anemic cases. The presence of other species of bacteria only in anemic and not in non-anemic samples may pinpoint a possible active competing role of these bacteria or preventing the absorption of iron-containing haem precursors. *P. pseudoalcaligenes* and *P. putrefaciens* (45 isolates), for example were detected only in anemic cases and not in volunteer control samples. The reciprocal relation obtained between the HB and ESR values, as well as the linear relation that was observed between ESR values and TVB counts in stool samples is of remarkable finding. Bacterial toxins are a well-known cause of high ESR values. This triagonal relation thought to be an indirect relation by Cotran *et al.* (1999) but when there is no clear clinical reason for anemia, this relation should be considered direct and important. In this study, for example, the lowest investigated figure of hemoglobin was HB 7.2, had ESR values of 110 after both 1 and 2 hours and found to contain TVB counts as high as 8298 x 10\(^6\) cfu/g of stool sample. The anemic case with HB value of 9.5 and ESR reading of 25, 50 after 1, 2 hours respectively, found to harbor TVB counts of only 40 x 10\(^6\) cfu/g of stool sample. The control case with HB value of 13 and ESR readings of 2, 4 after 1, 2 hours respectively, proved to have only 9 x 10\(^5\) cfu/g of stool sample. These results confirmed the discussed correlation.

The recorded, more or less far, two points from this relation in this study could be not due to the quantity of the bacteria but due to the quality of them. In the anemic case, for example, that had BH value of 7.8 and relatively low ESR reading of 18, 40 after 1, 2 hours respectively, showed almost the highest TVB count 31767 x 10\(^6\) cfu/g of stool sample. According to the previously explained relation, it was supposed to have ESR values more than 110. This sample had a wider range of the bacterial population than other studied samples; more than those detected in the stool sample of the case with HB value of 7.2, but lacking the presence of one important species, *A. punctata*. Many species of this genus have been long documented as human intestine-inhabitants (Agger *et al.*, 1985; Moyer 1987, Challapalli *et al.*, 1988; De la Morena *et al.*, 1993). Out of the identified bacteria in this study 6
species were detected only in stool samples of anemic cases; i.e. not detected in control stool samples. These 6 species namely; *P. putrefaciens*, *M. luteus*, *E. rhusiopathiae*, *B. megaterium*, *B. pumilus* and *B. coagulans*, are also noticed to found in low HB and high TVB counts. *E. rhusiopathiae* for example isolated among TVB counts of 1516 x 10^6 cfu/g from anemic case with HB of 9.5 g/dl. These six species were found sensitive to almost all the tested antibiotics; this indicates how easy their control is. Then the prevention of their suggested counterpart in competing for the iron compounds in normal human intake will diminish to a minimum and, in turn, the need for supplemental iron medication could be avoided. *C. equitium* was the only isolate that showed resistance to two; colistin sulfate and erythrocin, of the six tested antibiotics in this study, while, *C. pseudotuberculosis* was not.

**CONCLUSION**

This research addressed a new scope of an old problem. The study confirmed the hypothesized relation between the intestinal population of bacteria both, quantitative and qualitative, and the symptoms of iron deficiency anemia through the possible active competition mechanisms of bacteria on iron compounds as heme precursors. The study recommends the necessity of a culture and sensitivity test to be done for the iron-deficiency anemic patients in Egypt. Prescription of the appropriate antibiotic well then is effective in minimizing the need for supplemental iron medications.

**REFERENCES**


