Screening of lignocellulolytic fungi for hydrolyzation of lignocellulosic materials in paddy straw for bioethanol production

Mona Fati Syazwane Mohamed Ghazali1, Nur Ain Izati Mohd Zainudin1, Nor Azwady Abd Aziz2 and Muskhazli Mustafa**

1Department of Biology, Faculty of Science, Universiti Putra Malaysia, 43400 UPM Serdang, Selangor
Email: muskhazli@upm.edu.my

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ABSTRACT

Aims: Paddy straw is known to have lignocellulosic materials such as cellulose and hemicellulose which can be readily converted into fermentable sugar for production of bioethanol via simultaneous saccharification and fermentation (SSF). In order to produce ethanol competently, the degradation of biomass by cellulase and highly ethanol-producing microorganism in fermentation process are necessarily needed. However, there is lacking in cellulose degrading organism in producing adequate amount of lignocellulosic enzyme. Therefore, the screening and selection for the best fungi to hydrolyze the lignocellulosic materials as well as forming consortium between two species of fungi has become the main focus. Methodology and results: Thirteen strains of fast-growing fungi were tested qualitatively for cellulase (congo red staining) and polyphenol oxidase (Bavendamm test). All tested strains displayed lignocellulolytic fungi characteristics. The selection was narrowed down by quantitative assay on endoglucanase, exoglucanase, β-glucosidase and xylanase and the highest cellulases enzyme producer were T. asperellum B1581 (3.93 U/mL endoglucanase; 2.37 U/mL exoglucanase; 3.00 IU/mL β-glucosidase; 54.87 U/mL xylanase), followed by A. niger B2484 (5.60 U/mL endoglucanase; 1.08 U/mL exoglucanase; 1.57 IU/mL β-glucosidase; 56.85 U/mL xylanase). In compatibility test, both T. asperellum B1581 and A. niger B2484 were inoculated on the same petri dish for 4 days and the interaction showed by the two species was mutual intermingling. Conclusions, significance and impact of study: Both Trichoderma asperellum B1581 and Aspergillus niger B2484 produced the highest cellulase enzyme. Since both strains can co-exist and produce enzymes that complete each other, a fungal consortium was suggested to increase the yield of sugars in saccharification process.

Keywords: Cellulose, hemicellulose, consortium, qualitative assay, quantitative assay

INTRODUCTION

In December 2017, Food and Agriculture Organization (FAO) has increased its estimation of paddy production across the globe in the year 2017 by 2.9 million tonnes to 759.6 million tonnes and China remains the largest paddy producer in the Asia region (FAO, 2018). The increased demand of rice production has led to an increase amount of world’s paddy straw production, with an estimation of at least 73.1 million tons per annum (Wu et al., 2013). Paddy straw is the residual of rice production and has become one of the most commonly available lignocellulosic waste materials (Saini et al., 2015). Current disposal method by burning not only resulted in reducing the air quality (Rosmiza et al., 2014) but also wasting the potential of harvesting valuable carbohydrates for biofuel production as the paddy straw contains about 45-75% of complex carbohydrates (Fan et al., 2013). Bioethanol is a renewable energy source that have prospect in handling current world energy issues and the deteriorating quality of environment (Aditiya et al., 2016). Now, most technologies involve in biofuel production are focusing on turning lignocellulose biomass into transportation fuels (Bakker et al., 2013).

The first generations of biofuels are originated from crop plants, but they produced limited biofuel yields and threatening on food security (Aro, 2016). Alternative feedstock also known second-generation feedstock, are recovered from woody biomass which are more energy efficient, flexible and not competing with the human’s food resources (Havlík et al., 2011). An estimation of 442 billion liters of bioethanol can be produced from lignocellulosic waste material, which the amount is 16 times higher than the real amount of bioethanol produced globally (Sarkar et al., 2012).

**Corresponding author
The bioconversion of biomass into bioethanol involves several processes such as the pretreatment, saccharification and fermentation (Oberoi et al., 2012). Cellulolytic enzymes involve in hydrolysis of lignocellulosic materials (saccharification), which are commonly broken down by cellulolytic bacteria and fungi (Belal, 2013). The by-products of the saccharification process are generally reducing sugars including glucose. According to Okamoto et al. (2011), several filamentous fungi has been explored for their capability in producing ethanol from biomass, such as the genera Mucor, Rhizopus, Neurospora, Aspergillus, Fusarium, Monilia, and Trichoderma. Most of the Trichoderma sp. are fast-grower, produce large amount of conidia and variety of valuable enzymes such as cellulase (Khan et al., 2007). Cellulase is also known as cellulolytic enzymes, of which three classes are identified on the basis of the mode of the substrate specificities and enzymatic actions: endoglucanases (EC 3.2.1.4), exoglucanases (EC 3.2.1.74 and EC 3.2.1.91) and β-glucosidases (EC 3.2.1.21) (Teeri, 1997; Kuhad et al., 2016). Only with the synergism of the above three enzymes makes the complete hydrolysis of cellulose into glucose (Gupta et al., 2012). An efficient and cost-effective enzyme system should contain balanced activities of cellulases (both endo- and exo-glucanase), β-glucosidase, and xylanase, and the system should also have high titer of enzyme activities to balance the cost of ethanol production (Brijwani et al., 2010).

The simultaneous saccharification and fermentation (SSF) processes involves enzymatic saccharification of cellulose with fermentation of glucose to ethanol (Kusmiyati et al., 2018). It has become favorable process as it improved the yield of ethanol, via reducing the product inhibition excreted and also removes the need for separate reactors, which results in cost reduction and reduce the risk of contamination (Wang et al., 2013; Narra et al., 2015).

Regardless the fact that some of the fungal strains have the benefits of being lignocellulolytic and thermostable, most of these fungal strains failed to produce adequate amounts of lignocellulolytic enzymes, which are essentially required for efficient bioconversion process (Dashbun et al., 2009). According to Kausar et al. (2010), mixed cultures of consortium are able to stimulate colonization of the substrate by increasing the production of enzymes, withstand contamination by other microbes and results in high yield of sugars. However, the selection of suitable cellulase for hydrolyzation process of lignocellulosic materials is very difficult because each biomass has structural and enzymatic activities differences (Takano and Hoshino, 2018). Hence, finding appropriate lignocellulolytic fungi for degradation of paddy straw is necessarily needed. In order to address the foregoing issues, this study was commenced to screen filamentous fungi with an ability to secrete hydrolytic enzyme for substrate degradation and to test the compatibility between two different species of fungi in an attempt to produce better yield of sugars in saccharification process.

**MATERIALS AND METHODS**

**Collection of fungi**

Fast growing and molecularly identified fungi strains (Table 1) were obtained from Mycology Laboratory, Faculty of Science, Universiti Putra Malaysia and School of Biological Sciences, Universiti Sains Malaysia. All strains were grown on Potato Dextrose Agar (PDA) incubated at 28 ± 2 °C for 7 days.

<table>
<thead>
<tr>
<th>Fungi species</th>
<th>Strain</th>
<th>Source</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trichoderma asperellum</td>
<td>B1581</td>
<td>Soil</td>
<td>Sharifah Siti Maryam et al., 2016</td>
</tr>
<tr>
<td>Trichoderma asperellum</td>
<td>B1584</td>
<td>Soil</td>
<td>Sharifah Siti Maryam et al., 2016</td>
</tr>
<tr>
<td>Aspergillus niger</td>
<td>B2484</td>
<td>Vietnamese coriander</td>
<td>Nur Ain Izzati et al., 2017</td>
</tr>
<tr>
<td>Aspergillus niger</td>
<td>C2472</td>
<td>Winged bean</td>
<td>Nur Ain Izzati et al., 2017</td>
</tr>
<tr>
<td>Fusarium oxysporum</td>
<td>B633T</td>
<td>Tomato</td>
<td>Nur Baiti et al., 2016</td>
</tr>
<tr>
<td>Fusarium oxysporum</td>
<td>B645T</td>
<td>Tomato</td>
<td>Nur Baiti et al., 2016</td>
</tr>
<tr>
<td>Fusarium fujikuroi</td>
<td>688</td>
<td>Rice</td>
<td>-</td>
</tr>
<tr>
<td>Fusarium fujikuroi</td>
<td>4851</td>
<td>Rice</td>
<td>-</td>
</tr>
<tr>
<td>Fusarium fujikuroi</td>
<td>4872</td>
<td>Rice</td>
<td>-</td>
</tr>
<tr>
<td>Curvularia lunata</td>
<td>P1221</td>
<td>Rice</td>
<td>Nur Ain Izzati et al., 2019</td>
</tr>
<tr>
<td>Curvularia lunata</td>
<td>P1244</td>
<td>Rice</td>
<td>Nur Ain Izzati et al., 2019</td>
</tr>
<tr>
<td>Curvularia eragrostis</td>
<td>P1262</td>
<td>Rice</td>
<td>Nur Ain Izzati et al., 2019</td>
</tr>
<tr>
<td>Curvularia eragrostis</td>
<td>K872</td>
<td>Rice</td>
<td>Nur Ain Izzati et al., 2019</td>
</tr>
</tbody>
</table>

**Qualitative assays of Lignocellulolytic activity**

**Cellulose degradation**

A 5-mm mycelial disc was cut using cork borer from the edge of fungal culture (7 days old) and transferred to a prepared media (carboxymethylcellulose sodium salt 20.0 g/L; agar 15.0 g/L) before incubated at 28 ± 2 °C for 7 days as proposed by Mukhlis et al. (2013). The plate was stained with 1 mg/mL Congo red solution and formation of halo ring (clear zonation) around the fungal colony indicates the cellulase activity of the fungal (Kausar et al., 2010). The diameter of halo ring was measured and enzymatic index (EI) were calculated based on formula of

\[ EI = \frac{D_h - D_s}{D_h} \times 100 \]

where, \( D_h \) is the diameter of the halo ring and \( D_s \) is the diameter of the fungal colony.
Enzymatic Index = Diameter of Halo Ring / Diameter of Growth Colony (Floresco et al., 2012).

**Bavendamm test for laccase detection**

Presence of polyphenol oxidase (PPO) indicates the existence of laccase. A disc of fungal hyphae was placed at the center of the prepared media (Tannic acid 5.0 g/L; Malt extract agar 15.0 g/L; Difco Agar 20.0 g/L) and incubated for 7 days a 28 ± 2 °C in dark condition to determine the presence of polyphenol oxidase (PPO) activity (Thomann et al., 2002). PPO was used to indicate laccase activity on lignin and the diameter of brown pigmentation formed around fungal colony was measured generally after five days.

**Quantitative assays**

The quantitative assays were also carried out to determine the activity of cellularase and hemicellulase enzymes in order to narrowing down to the best fungi producing hydrolytic enzyme. An amount of 1 x 10⁶ spore/mL were cultured in basal medium (NH₄)₂SO₄ 1.4 g/L; KH₂PO₄ 2.0 g/L; CaCl₂ 0.3 g/L; MgSO₄·7H₂O 0.3 g/L; CoCl₂ 2.0 g/L) added with 1 mL of trace element (MnSO₄·H₂O 1.56 g/L; FeSO₄·7H₂O 5.0 g/L; ZnSO₄·7H₂O 1.4 g/L) for 7 days at 150 rpm and 30 ± 1 °C (Ja’afaru, 2013) before crude culture was collected for quantitative assay.

**Endoglucanase, exoglucanase and xylanase assays**

The cellulase and hemicellulase enzymes were assayed using 3, 5-dinitrosalicylic acid (DNS) method (Miller, 1959). Carboxymethyl-cellulose (CMC), Avicel® PH-101 and xylan from corn were used as substrate for endoglucanase, exoglucanase and xylanase assay, respectively. An amount of 0.09 mL substrate-buffer (1% (w/v) substrate; 0.05 M sodium acetate pH 4.8) was mixed with 0.01 mL of crude enzyme and incubated at 50 ± 1 °C for 30 min before 0.1 mL DNS were added to stop the reaction (Das et al., 2012; Ja’afaru, 2013). The absorbance was read at 540 nm and the amounts of sugar were determined using glucose standard (for endoglucanase and exoglucanase) and xyllose standard (for xylanase). One Unit (U) of enzyme activity was defined as the release of 1 μmol of reducing sugar per min.

**β-glucosidase assay**

A mixture of 0.025 mL of substrate-buffer (1% (w/v) 4-nitrophenol β-D glucopyranoside; 0.05 M sodium acetate pH 4.8) and 0.025 mL of crude enzyme were incubated at 50 ± 1 °C for 15 min before added with 0.05 mL 1 M Na₂CO₃ to stop the reaction (Jeya et al., 2010). The absorbance was read at 400 nm and the amount of p-nitrophenol (pNP) released was quantified based on pNP standard. One International Unit (IU) of β-glucosidase activity was expressed as the amount of enzyme required to release 1 μmole of pNP per min under the assay conditions.

**Compatibility test**

Seven days old fungal mycelial discs of selected fungi were place on the same PDA with a gap of 4 cm from each other (Kausar et al., 2010). The plate was incubated at 28 ± 1 °C. Their growth rate and pattern were observed everyday till full grown; which the fungi mycelia fully occupied the surface of the agar (approximately 5 days). Mode of interaction between the filamentous fungi was observed and identified using a key reference by Mohammad et al. (2011) as shown in Table 2.

<table>
<thead>
<tr>
<th>No.</th>
<th>Interaction</th>
<th>Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Mutual intermingling</td>
<td>Both fungi successfully grow into each other without any macroscopic indication of interconnections.</td>
</tr>
<tr>
<td>2</td>
<td>Partial intermingling</td>
<td>One of the funguses grows into the other fungus either above/below without forming any inhibition zone.</td>
</tr>
<tr>
<td>3</td>
<td>Invasion/replacement</td>
<td>One mycelium propagates into the other and begins to ingest the weaker one before colonize it.</td>
</tr>
<tr>
<td>4</td>
<td>Inhibition at contact point</td>
<td>The fungi almost make contact with each other with a fine segregation line (1 to 2 mm) between the two colonies.</td>
</tr>
<tr>
<td>5</td>
<td>Inhibition at distance</td>
<td>Inhibition at a distance of more than 2 mm</td>
</tr>
</tbody>
</table>

**RESULTS AND DISCUSSION**

Microorganisms in which secretes cellulase have a major role in the environment, as they have decaying ability of biomass, and hence creating a crucial relation in the carbon cycle (Zhang et al., 2006; Damaso et al., 2012). Many studies have been highlighted on fungi producing cellulases as they are abundant, easy to extract and some of the fungal cellulases have been used widely used for commercials (Liang et al., 2014). All thirteen strains of molecularly identified fungi (Table 1) were selected based on their origin and ability as fast-growing fungi. However, their lignocellulolytic characteristics have yet to be tested.

In early screening, qualitative assays were performed to screen the quality of cellulase and laccase secreted by the fungi. In the qualitative screening, all carboxymethyl-cellulose (CMC) was used to raise the molecular disorder of the densely arranged cellulotic network and to uncover the cellulose chains buried within the microfibris (Arantes and Saddler, 2010). Meanwhile the inefficiency of Congo red dye to bind with mono or disaccharides leading to a visible clear zone formation known as halo ring (Gohel et al.)
and relatively the diameter of halo ring produced is proportionate with the level of cellulase activity. An enzymatic index of ≥ 0.5 is considered to be the best producers of cellulase enzyme. As shown in Table 2, out of 13 strains assayed, only 8 were found to have index more than 0.5. The largest enzymatic index (EI) was produced by Trichoderma asperellum B1584 at the value of 0.88. While the smallest EI value was produced by Curvularia eragrostidis P1262 with 0.25. However, there was no significant difference between all the samples tested.

Theoretically in Bavendamm test, utilization of phenolic compounds such as gallic acid or tannic acid in nutrient agar by the white rot fungi produce a deeply brown colored zone around the mycelium and this zone has become an indicator for the quality of the fungi polyphenol oxidase (PPO) activity (Mushimiyimana and Tallapragada, 2014). The ratio of brown pigmentation to growth colony was measured (Table 3) and the largest ratio produced was 1.60 by C. eragrostidis P1262 but it was not significantly different from other samples. All Curvularia sp. showed their ability to degrade lignin with the presence of polyphenol oxidase (PPO) but somehow failed to produce a good response in Congo red assay. The quality of cellulase has become the key of fungi selection as the successfullness of the whole bioconversion process mainly depends on the sources and production of cellulolytic enzyme as well as the optimal conditions for catalytic activity (Ram et al., 2014). Therefore, both C. lunata and C. eragrostidis has been removed from list for the quantitative assays. To narrow down the selection and precisely measures the cellulase and hemicellulase activity, the quantitative assays were done as a secondary screening for final selection.

Trichoderma asperellum B1581 came out as the best producer of exoglucanase and β-glucosidase at 2.37 ± 0.34 U/mL and 3.00 ± 0.15 U/mL, respectively (Table 4). On the other hand, Aspergillus niger B2484 had the highest endoglucanase (5.60 ± 0.43 U/mL) and xylanase (56.85 ± 2.75 U/mL) and considerably high amount of exoglucanase as well as β-glucosidase (1.08 ± 0.17 U/mL; 1.57 ± 0.02 U/mL); thus making them as one of best fungi producing cellulase along with T. asperellum B1581.

The high production of exoglucanase; also known as cellubiohydrolase (CBH) is essential as they represent 60% of the enzyme cocktail population; primary exocellulase and responsible to degrades cellulose into cellbiose (Brady et al., 2015). According to Wang et al. (2012), the optimized ratios of cellulolitic-enzymes for the best saccharification of crystalline cellulose is 60% CBH I: 20% CBH II: 20% endoglucanase. β-glucosidase plays a significant role in bioethanol production to eliminate cellbiose inhibition (Whang et al., 2012) and it has become a conundrum in producing bioethanol.

Table 3: Qualitative assessment on fungi strains for selection of the best fungi producing hydrolytic enzymes.

<table>
<thead>
<tr>
<th>Fungi Strain</th>
<th>Cellulase (EI)**</th>
<th>Laccase (ratio)</th>
</tr>
</thead>
<tbody>
<tr>
<td>A. niger B2484</td>
<td>0.78bc</td>
<td>1.01±</td>
</tr>
<tr>
<td>A. niger C2472</td>
<td>0.64abc</td>
<td>1.00±</td>
</tr>
<tr>
<td>C. eragrostidis P1262</td>
<td>0.25abc</td>
<td>1.60±</td>
</tr>
<tr>
<td>C. eragrostidis K872</td>
<td>0.26abc</td>
<td>1.55±</td>
</tr>
<tr>
<td>C. lunata P1221</td>
<td>0.38c</td>
<td>1.43cd</td>
</tr>
<tr>
<td>C. lunata P1244</td>
<td>0.36bc</td>
<td>1.45bc</td>
</tr>
<tr>
<td>F. fujikuroi 688</td>
<td>0.49ab</td>
<td>1.32bcde</td>
</tr>
<tr>
<td>F. fujikuroi 4872</td>
<td>0.58bc</td>
<td>1.31bcde</td>
</tr>
<tr>
<td>F. fujikuroi 4851</td>
<td>0.58bc</td>
<td>1.18cdef</td>
</tr>
<tr>
<td>F. oxyporum B633T</td>
<td>0.61abc</td>
<td>1.13cdef</td>
</tr>
<tr>
<td>F. oxyporum B645T</td>
<td>0.62abc</td>
<td>1.10cdef</td>
</tr>
<tr>
<td>T. asperellum B1581</td>
<td>0.68abc</td>
<td>1.29cdef</td>
</tr>
<tr>
<td>T. asperellum B1584</td>
<td>0.88a</td>
<td>1.35cdef</td>
</tr>
</tbody>
</table>

Values are means of three replicates with ±SD.
*Means in each column with same superscript letter are not significantly different amongst themselves when Tukey test were used at 5% significance level
** Enzymatic Index (EI) is a measure for cellulase activity

Table 4: Qualitative and quantitative assessment on fungi strains for selection of the best fungi producing hydrolytic enzymes.

<table>
<thead>
<tr>
<th>Fungi Strain</th>
<th>Endoglucanase (U/mL)</th>
<th>Exoglucanase (U/mL)</th>
<th>β-glucosidase (IU/mL)</th>
<th>Xylanase (U/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>A. niger B2484</td>
<td>5.60 ± 0.43±</td>
<td>1.08 ± 0.17cdef</td>
<td>1.57 ± 0.02ab</td>
<td>56.85 ± 2.75a</td>
</tr>
<tr>
<td>A. niger C2472</td>
<td>1.83 ± 0.15±</td>
<td>1.75 ± 0.20abc</td>
<td>0.05 ± 0.03c</td>
<td>27.77 ± 2.69d</td>
</tr>
<tr>
<td>F. fujikuroi 688</td>
<td>0.98 ± 0.01±</td>
<td>1.01 ± 0.14abcdef</td>
<td>0.09 ± 0.04c</td>
<td>56.00 ± 2.82a</td>
</tr>
<tr>
<td>F. fujikuroi 4872</td>
<td>2.14 ± 0.09±</td>
<td>0.25 ± 0.14±</td>
<td>0.03 ± 0.02c</td>
<td>37.69 ± 1.39cd</td>
</tr>
<tr>
<td>F. fujikuroi 4851</td>
<td>0.95 ± 0.04±</td>
<td>1.17 ± 0.16bcde</td>
<td>0</td>
<td>32.04 ± 9.19cd</td>
</tr>
<tr>
<td>F. oxyporum B633T</td>
<td>1.10 ± 0.09±</td>
<td>0.95 ± 0.23cdef</td>
<td>0</td>
<td>43.41 ± 6.92abcd</td>
</tr>
<tr>
<td>F. oxyporum B645T</td>
<td>0.93 ± 0.06±</td>
<td>2.04 ± 0.53ab</td>
<td>0</td>
<td>42.28 ± 2.06bcd</td>
</tr>
<tr>
<td>T. asperellum B1581</td>
<td>3.93 ± 0.05±</td>
<td>2.37 ± 0.34a</td>
<td>3.00 ± 0.15a</td>
<td>54.87 ± 8.23ab</td>
</tr>
<tr>
<td>T. asperellum B1584</td>
<td>1.69 ± 0.10±</td>
<td>2.00 ± 0.49ab</td>
<td>0.03 ± 0.02c</td>
<td>34.55 ± 4.69cd</td>
</tr>
</tbody>
</table>

Values are means of three replicates with ±SD.
*Means in each column with same superscript letter are not significantly different amongst themselves when Tukey test were used at 5% significance level.
One of the major challenges in producing bioethanol is to search for a glucose tolerant β-glucosidase as they are the key component present in cellulase to complete the final step during cellulose hydrolysis and conversion process of cellulose into glucose (Singhania et al., 2012). Currently, commercial cellulase was prepared using T. reesei due to its high activities of both exoglucanase and endoglucanase (Sohail et al., 2009). However, low levels of β-glucosidases in T. reesei have been complemented by mixing β-glucosidase from A. niger in T. reesei cellulolytic cocktail (Del Pozo et al., 2012; Rani et al., 2014). As in this study, combining these two species is expected to produce higher glucose yield for bioethanol production since T. asperellum B1581 obtained high exoglucanase and β-glucosidase activities while A. niger B2484 attained high endoglucanase and xylanase activities. In relation to qualitative assays, it was found that the diameter of clearance zone (in Congo red) exhibited almost a linear relationship with the enzyme activity.

Microbial interactions are usually referred as hostile, but they can also be supportive, related with a metabolic change of each partner and cause a modification in the arrangement of produced bioactive molecules (Benoit-Gelber et al., 2017). Consortia are a favorable method that could help to understand the synergistic effects between mixed enzymes for the biodegradation of lignocellulosic waste (Taha et al., 2015). The formation of consortia could be beneficial by eliminating problems such as incomplete synergistic enzymes and inhibition of end-product (Sheng et al., 2016). Through consortia, cellulase production was enhanced and the composition of cellulolytic enzymes was improved (Wen et al., 2005; Fang et al., 2013). Commercially, enzyme sets of T. reesei for cellulose degradation are often complemented with β-glucosidases from A. niger by combining T. reesei Cellobiast® and A. niger’s β-glucosidase Novozym188® (Brink et al., 2014). A study by Liu et al. (2017) shows Zygomonas grass could be utilized consortia of T. reesei and A. niger to produce glucose for bioethanol production by Zygomonas mobilis. However, according to Silva (2016), major obstacle is the compatibility since formation of consortia involves interrelationship in managing the interaction, waste, space and resource to achieve optimum mutualism. Therefore prior to any fungi application as consortia for production, the compatibility test for fungal consortia is essential. In compatibility test, both T. asperellum B1581 and A. niger B2484 were grown on a same plate and in-vitro interaction between T. asperellum B1581 and A. niger B2484 after 5 days of incubation shown no negative interaction (Figure 1). The interaction was mutual intermingling growth where the fungus being observed is growing into the opposed fungus either above or below its colony. There was no visible boundary formed which indicates that both T. asperellum B1581 and A. niger B2484 can live together in the same culture. Since both of the strains can mutually co-exist, it was suggest mixing the cellulolytic cocktails of different fungi strains could enhance the end-product after saccharification process (Kalyani et al., 2013).

Figure 1: shows the interaction between T. asperellum B1581 and A. niger B2484 after 5 days of incubation. (a) Front view (b) Rear view

CONCLUSION

Out of thirteen strains of filamentous fungi which were qualitatively and quantitatively assayed, the best two fungi strain were T. asperellum B1581 produces the highest exoglucanase and β-glucosidase (2.37 ± 0.34 U/mL; 3.00 ± 0.15 IU/mL respectively), while A. niger B2484 produces the highest endoglucanase and xylanase (5.60 ± 0.43 U/mL; 56.85 ± 2.75 U/mL respectively). Both of the fungi strain also showed their ability to formed mutual intermingling consortia in the in-vitro interaction of compatibility test. Since both of the strain can co-exist and produces enzymes that complete each other, the fungal consortium between T. asperellum B1581 and A. niger B2484 was suggested to increase the yield of sugars in saccharification process.

ACKNOWLEDGEMENTS

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cellulase production by Paenibacillus terrae ME27-1. 


